

Gas Phase IR spectroscopy:

towards large biomolecules & far-IR wavelengths

Anouk M. Rijs

Winterschool Han-sur-Lesse 2012, 10-14 December 2012

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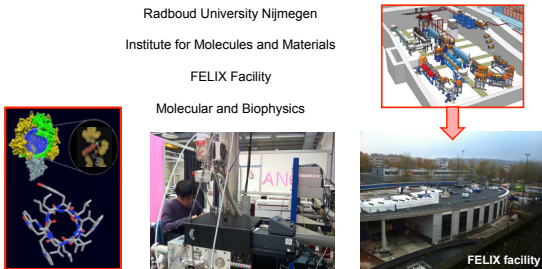
dr. Anouk Rijs

Radboud University Nijmegen

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Molecular and Biophysics



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Layout topics to discuss

Introduction

- Why, o why?
- What & how?

Experimental methods

- Cooling methods
 - Why cooling
 - Supersonic expansions
 - Collisional cooling
- Molecule production → larger & larger
 - heating
 - Laser desorption
 - Molecular beams & laser desorption

Spectroscopic Techniques

- UV + mass selective detection
- IR
- Table top IR vs Free electron lasers
- Combined methods:
 - IR-UV, UV-UV
 - Combination with VUV sources

Applications

- Molecular motors: functioning of rotaxanes
- Biomolecules
 - Local environment
 - Biological environment
 - Larger and larger....

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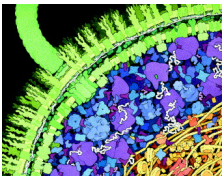
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Biomolecules in the gas phase: Why??

The biological medium is very complex

- Molecular crowding



cross-section of part of E.coli cell

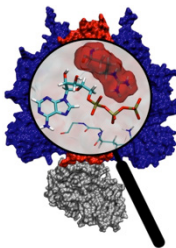
Questions that arise...

- Who does what?
- Does it do it by itself?
- Can the environment affect the molecule's role or functioning?
- What can go wrong?
 - car engine - living system
 - gears - isolated molecule
- What are the minimum requirements to do the job?

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Biomolecules in the gas phase: Why??



If we want to **understand the details** how the whole thing function we need to **understand its components**

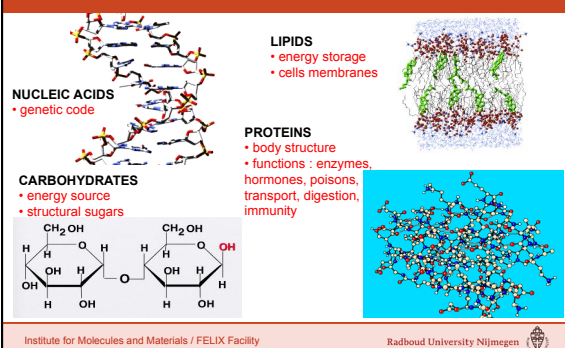
The **best environment** to study intrinsic properties of a molecular system* is **in vacuo** (no interactions, no collisions)

*(of limited size)

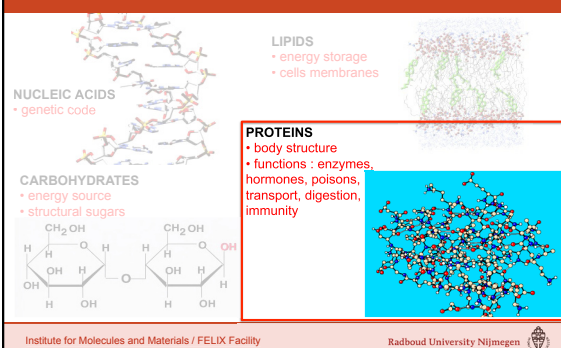
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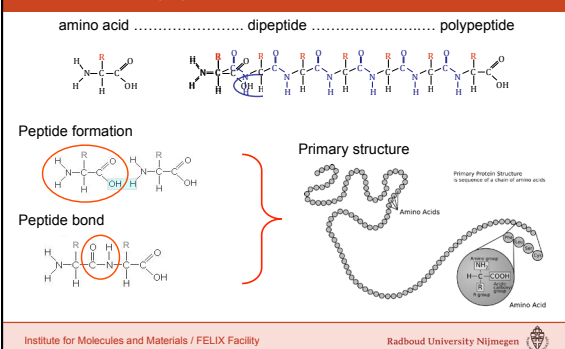
The four families of biomolecules



The four families of biomolecules



The structure of peptides



3D structure and function

Primary structure
linear sequence of amino-acids

1 KIPNFVPGK CASVDRLKW AEQTPNRSY AGWYGFALT
51 VRNEYSFDK QFVKSTGA YDGNLKRNG KLYPNFGEP HLSIDYENSF
101 AAPVLETD YSNVACLYSC IDNFQYHSD PSFSPRSAN LADQYKICE
151 AAPKNINVDI TRPVKTVQGS SCOPYDTQKTV

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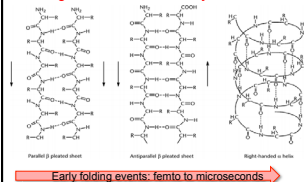
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Secondary structure
local organization stabilized by H bonds



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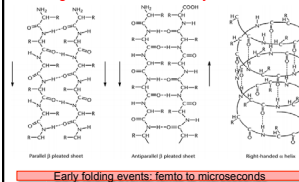
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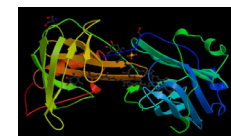
Secondary structure
local organization stabilized by H bonds



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Tertiary structure
3 dimensional form "native protein"



5 helices
9 beta strands
22 reverse turns
(3 disulfide bonds)

The function can only be carried out if the protein is in its native structure

folding in 3D structure: ~ 1 sec.

Protein folding & protein dynamics

What is the origin of irreversibility in nature?

Most biomolecules can be synthesized in various shapes. Yet in organisms, amino acids are always left-handed, and sugars are always right-handed. The origin of this preference remains a mystery.

How does protein folding occur?

Can we predict how proteins will fold? Out of a huge array, a protein picks one in just tens of milliseconds. The same task takes 10 years of computer time.

How does protein folding occur?

It has been hard enough counting genes. Proteins can be specified in different ways and described with numbers. Local properties and global properties are both important.

More the genome, find their functions?

Proteins are at the core of how life works. It is precise.

1 JULY 2005 VOL 317

2009 Science

Protein Dynamics

Big scientific issues: Still a lot what we don't know.....

Focus

Protein dynamics

Focus Issue: March 2012 Volume 4, No 3

Editorial Perspectives

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Understanding conformations

Nobel prize 1972: Anfinsen, Moore and Stein

amino-acid sequence + environment = one unique native structure

"for his work on ribonuclease, especially concerning the connection between the amino acid sequence and the biologically active conformation"

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Methods to study : static

X-ray crystallography : myoglobin 1959

Nuclear magnetic resonance : 1984

10 % of 100,000 human proteins

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Methods to study : dynamic

real time NMR → slow dynamics

Time-resolved methods → fast dynamics

Theoretical studies (both)

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Methods to study : conformation selective

diseases induced by misfolding:

Synthesis → Misfolded → Disordered aggregate → Intermediate → Native → Oligomer → Fibril → Crystal

Disordered aggregate → Degraded fragments → Degraded fragments → Degraded fragments

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conformation selective processes + local information

Flexibility and function

Biomolecular motors

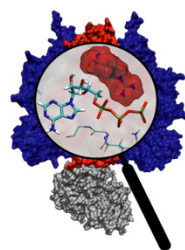
⇒ Biomolecular motion induced by conformational changes at the active site

⇒ Initiated by ATP association and hydrolysis

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Biomolecules in the gas phase: Why??



If we want to **understand the details** how the whole thing function we need to **understand its components**

The **best environment** to study **intrinsic properties** of a molecular system* is **in vacuo** (no interactions, no collisions)

*(of limited size)

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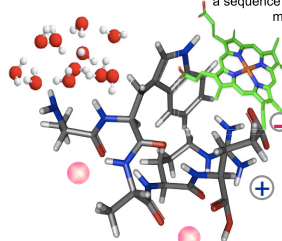
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Methods to study : conformation selective

HYPOTHESIS

a complex biological process

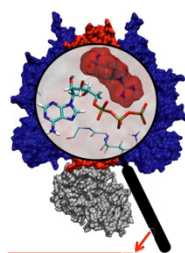
= a sequence of unit processes at the molecular level



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Biomolecules in the gas phase: What can we learn??



If we want to **understand the details** how the whole thing function we need to **understand its components**

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Conformation selective & local information

Who does what

Influence local and biological environment

IR spectroscopy

*(of limited size)

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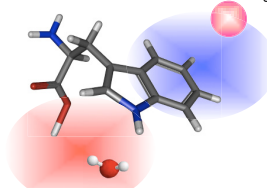
Conformation selective spectroscopic techniques

methods

Gas phase spectroscopy
Molecular beams = neutral molecules
Mass spectrometry (neutrals/ions)
Electronic (UV) spectroscopy
Vibrational (IR) spectroscopy

objects

isolated biomolecules
biomolecular interactions
- neutrals vs local charges
- water
- metals
- ligands



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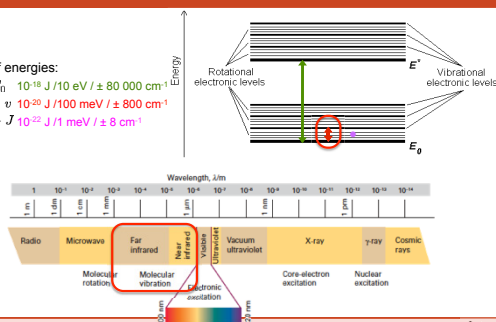
IR Spectroscopy

Orders of energies:

$$E_1 \leftarrow E_0 \quad 10^{-16} \text{ J} / 10 \text{ eV} / \pm 80\,000 \text{ cm}^{-1}$$

$$v + 1 \leftarrow v \quad 10^{-20} \text{ J} / 100 \text{ meV} / \pm 800 \text{ cm}^{-1}$$

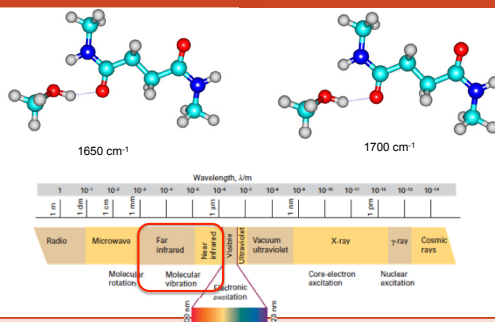
$$J + 1 \leftarrow J \quad 10^{-22} \text{ J} / 1 \text{ meV} / \pm 8 \text{ cm}^{-1}$$



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IR Spectroscopy



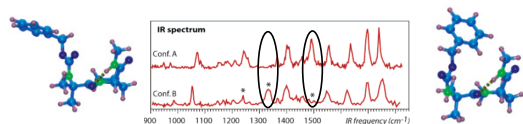
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Example

- direct view on **hydrogen bond interactions** (shift in IR frequency)

⇒ - provides detailed structural information



two conformations of a capped di-peptide distinguished by gas phase IR-UV spectroscopy

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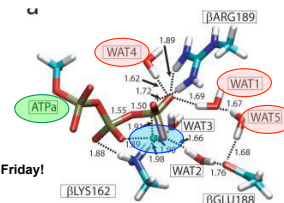
What can gas phase IR spectroscopy tell?

- direct view on **hydrogen bond interactions** (shift in IR frequency)

- provides detailed structural information

⇒ - influence of local and biological environment

- local: neighbouring amino acids
- biological:
 - water
 - metal ions /atoms
 - other molecules



Discuss in more detail on Thursday / Friday!

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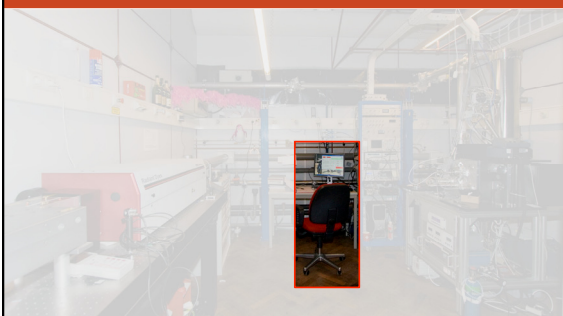
Applications

- Molecular motors: functioning of rotaxanes
- Biomolecules
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theoreticians toolbox



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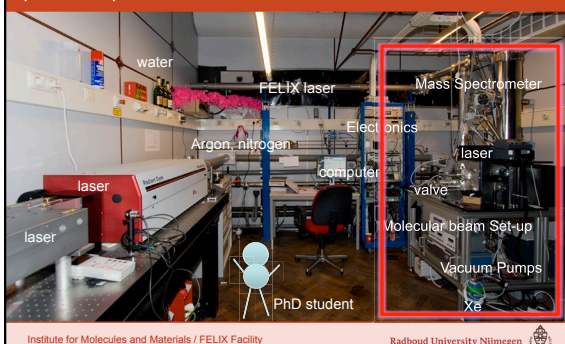
Spectroscopist's toolbox



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Spectroscopists toolbox



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Cooling molecules

As you know...

Biological processes take place at room temperature or in our case at 37°C

Question:

Why do we -spectroscopists- want to cool down molecules (or ions) to temperatures down to a few Kelvin??

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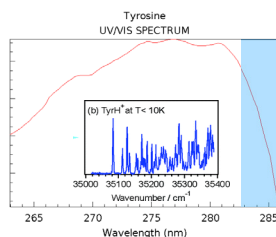
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JACS 128, p. 2816, 2006

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Answer:

To be able to analyze the spectra !!

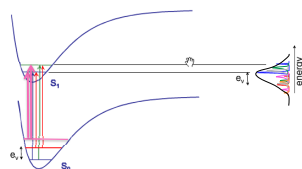
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Cooling molecules

How does cooling simplify spectra?



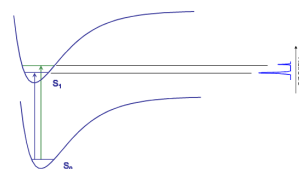
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Cooling molecules

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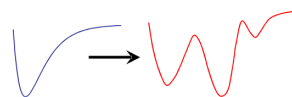
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Cooling molecules

Besides simplifying the spectra, what are the effects of cooling of large biomolecules



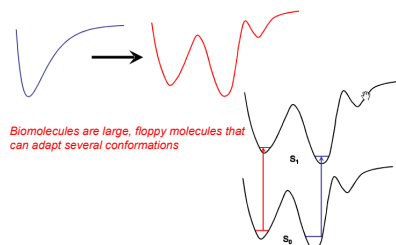
Biomolecules are large, floppy molecules that can adapt several conformations

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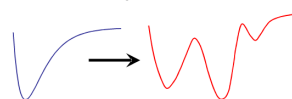
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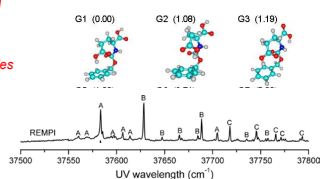
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Biomolecules are large, floppy molecules that can adapt several conformations

UV-UV hole-burning spectroscopy to locate origins of conformers



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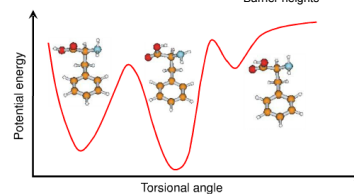
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Cooling molecules

Example: phenylalanine

Considering pro vs pre expansion conditions

- Average internal energy prior to expansion
- Cooling rate
- Relative energy of minima
- Barrier heights

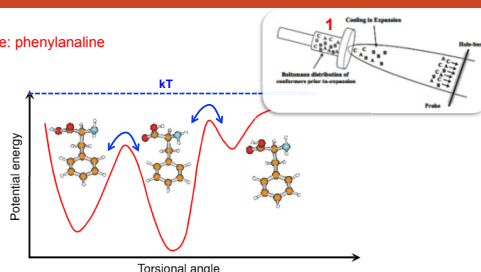


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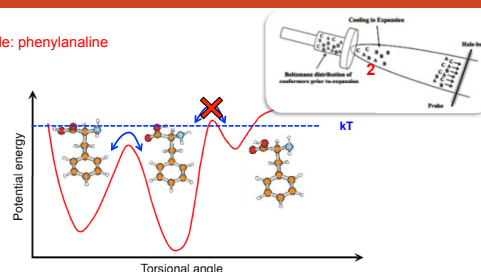


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Cooling molecules

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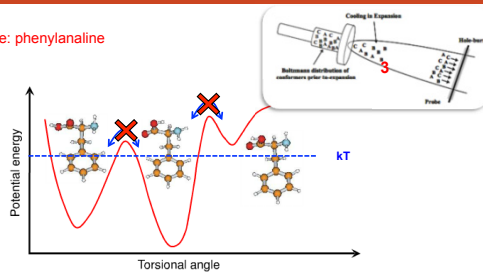


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Cooling molecules

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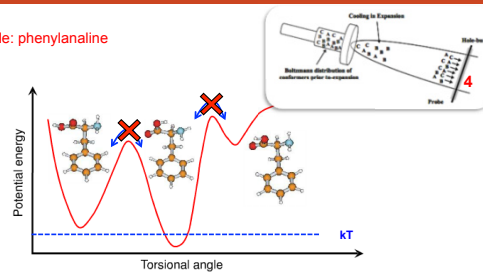


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Cooling molecules in a fast refrigerator ?!

Example: phenylalanine



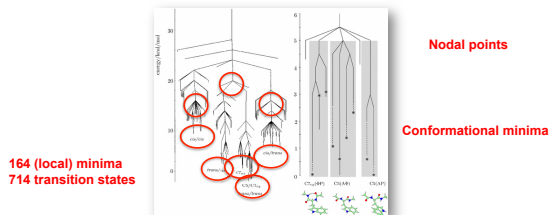
Cooling fast = population pre-expansion = pre-expansion

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Cooling molecules

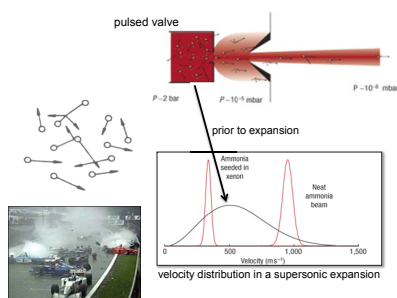
- When one studies cold molecules, it may or may not be the same systems as the ones at room temperature
- Depending on the cooling rate, we can end up with different conformational distributions



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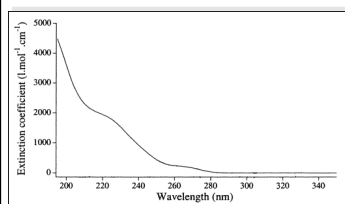
Supersonic expansion as a fast refrigerator



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Cold & Isolated Molecules



Thermal energy
→ molecules in different energy levels

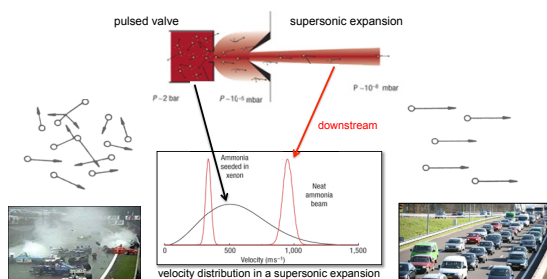
Intermolecular interactions
→ energy levels for each molecule different

F. Zerbetto et al., *J. Am. Chem. Soc.* 102, 149 (2002)

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Supersonic expansion as a fast refrigerator

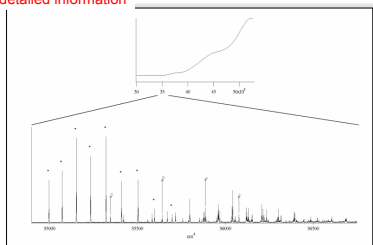


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Cold & Isolated Molecules

Unveiling detailed information

F. Zerbetto et al., *J. Am. Chem. Soc.* 102, 149 (2002)

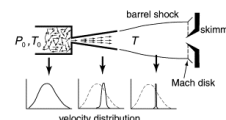
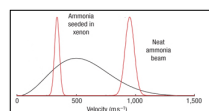
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Supersonic Expansion as a fast refrigerator

Supersonic expansions

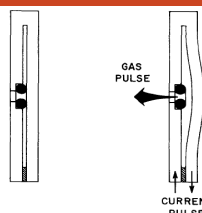
- Internal energy is converted to kinetic energy
 - Nearly monokinetic beam
 - Cooling of internal degrees of freedom
 - Carrier gas acts as a cooling bath for impurities (= molecules of interest)
- Zone of silence
 - No interference with background gas
 - Molecules studied in collision free regime
- High collision rate
 - Formation and stabilization of weakly bound complexes
 - Water clusters, metal ions, etc



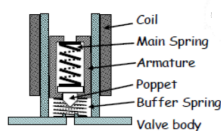
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Used types of valves



Precursor to the present-day *Jordan valve*: based on the principle of magnetic repulsion between two metal strips through which a large opposing current (3-5 kA) was pulsed.

W. R. Gentry and C. F. Giese, *Rev. Sci. Instrum.* 49, 595 (1978)

General Valve

magnetically activated plunger valve

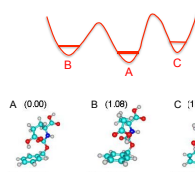
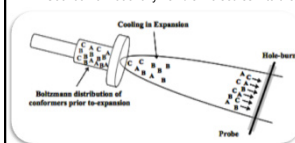
L. Abad, D. Bermejo, V. J. Herrero, J. Santos, and I. Tanarro, *Rev. Sci. Instrum.* 66, 3026 (1995)

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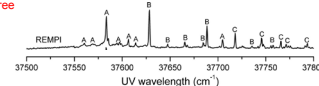
Supersonic Expansion applied to flexible biomolecules

Cool conformationally flexible molecules into the zero-point levels of low energy conformations



Spectroscopic challenge: Record the IR and UV spectra of single conformations! Free from interference from one another!

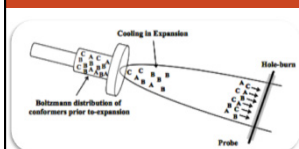
IR-UV ion dip spectroscopy



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Biomolecules in the gas phase: heating



Heating:

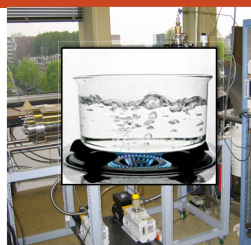
- volatile
- stable
- up to 300 C

- limited

- fragmentation?

From the 20 natural amino acids, only one (maybe two) possible to study via heating

Laser desorption

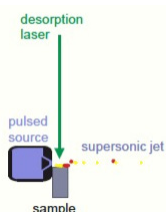


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Involatile biomolecules

Laser desorption combined with jet cooling

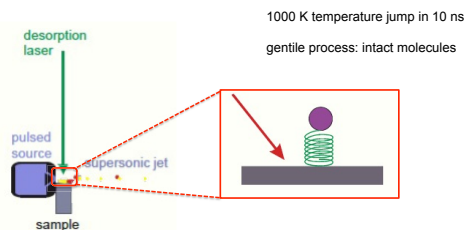


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= Laser desorption

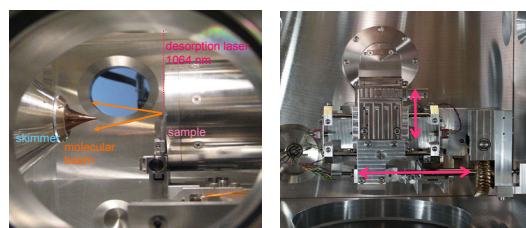
Laser desorption combined with jet cooling



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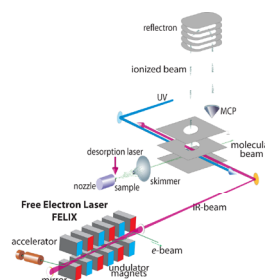
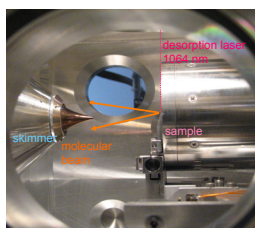
Experimental Method: molecular beam + laser desorption

laser desorption:
Creation gaseous molecules

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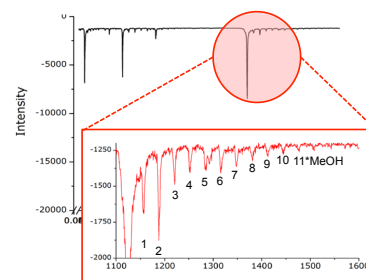
Experimental Method: molecular beam + laser desorption

laser desorption:
Creation gaseous molecules

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Controlled addition of the environment: solvent

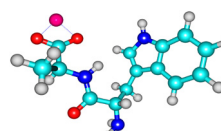


neutral rotaxanes

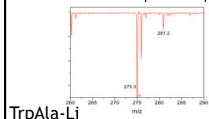
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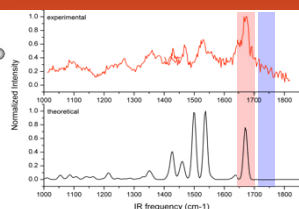
Controlled addition of the environment- metal ions

**Mass spectra:**

Metal ion has replaced a proton



TrpAla-Li

**IR-UV spectra:**
Carboxylic C=O is missing
↳ Gas phase salt

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Layout

Introduction

- Why, o why?
- What & how?

Experimental methods

- Cooling methods
 - Why cooling
 - Supersonic expansions
 - Collisional cooling
- Molecule production → larger & larger
 - heating
 - Laser desorption
 - Molecular beams & laser desorption

Spectroscopic Techniques

- History
- Use of lasers
- Experimental approach
 - UV, IR spectroscopy
- Free electron lasers
- Other methods:
 - IR-UV, UV-UV
 - Combination with VUV sources

Applications

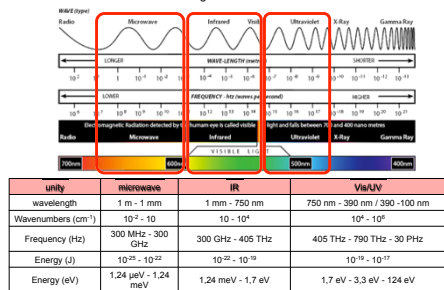
- Molecular motors: functioning of rotaxanes
- Biomolecules
 - Local environment
 - Biological environment
 - Larger and larger

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Spectroscopy

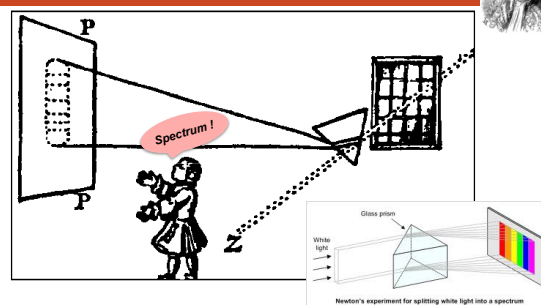
Interaction of molecules / atoms with electromagnetic radiation



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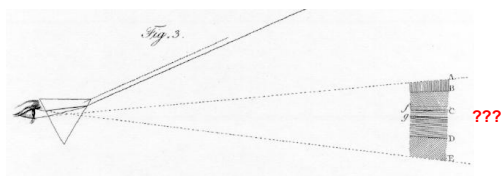
History of Spectroscopy: Newton – the year 1666



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History of Spectroscopy: Wollaston – the year 1802

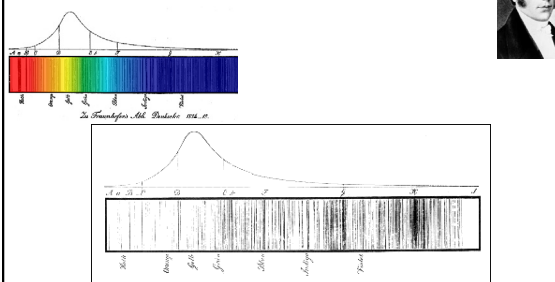


First observation of dark lines in the solar spectrum

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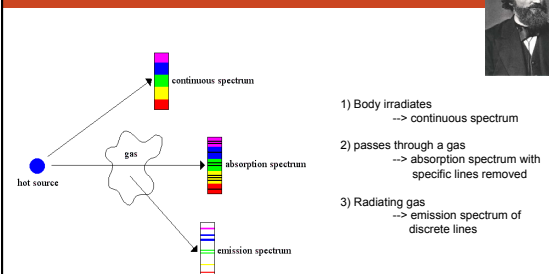
History of Spectroscopy: Fraunhofer – the year 1814



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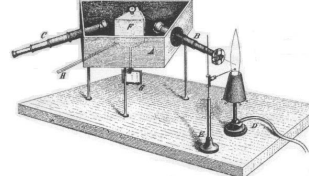
History of Spectroscopy: Kirchhoff's law - 1859



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History of Spectroscopy: Kirchhoff & Bunsen - 1860



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Spectroscopy in 2012

the lab ...

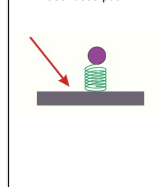


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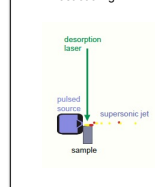
Modern experimental approach

Laser desorption



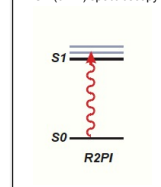
✓ (1) Pick them up

Jet cooling



✓ (2) Put them together

UV (or IR) spectroscopy

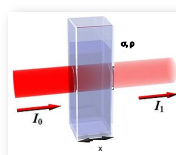


(3) Hold them up against the light

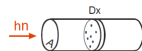
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Classical spectroscopy



Beer's law



$$\begin{aligned}\Delta I &= -\frac{\sigma}{A} N I \\ &= -\sigma \frac{N}{V} \Delta x \cdot I \\ &= -\sigma \frac{N}{V} \Delta x \cdot I \\ &= -\sigma \rho \Delta x \cdot I \\ \rightarrow \frac{\Delta I}{\Delta x} &= \frac{dI}{dx} = -\sigma \rho I\end{aligned}$$

$$\rightarrow I(x) = I_0 e^{-\sigma \rho x}$$

$$\frac{I}{I_0} = e^{-\sigma \rho x}$$

but:

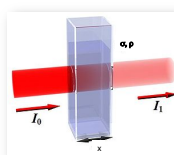
$$\text{NB: } \sigma [\text{cm}^2] \cdot \rho \left[\frac{1}{\text{cm}^3} \right] \cdot x [\text{cm}]$$

"Classical" spectroscopy: Requires optical depth (10^{-3})
→ selected experiments

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Restrictions



almost every lab has an absorption spectrometer

- identification
- structure
- geometry
- quantitative analysis
-

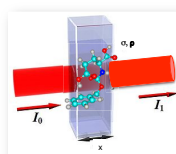
but:

"Classical" spectroscopy: Requires optical depth (10^{-3})
→ selected experiments

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Restrictions

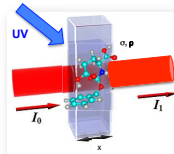


$$I_1 \approx 0 \rightarrow I_1 \approx I_0, \text{ no absorption signal visible}$$

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Action Spectroscopy



Dissociation
Fragmentation
Ionization
Electron Loss
....

main applications:

gas phase studies
molecules, clusters, complexes
neutral, ionic species
molecular beams, ion traps

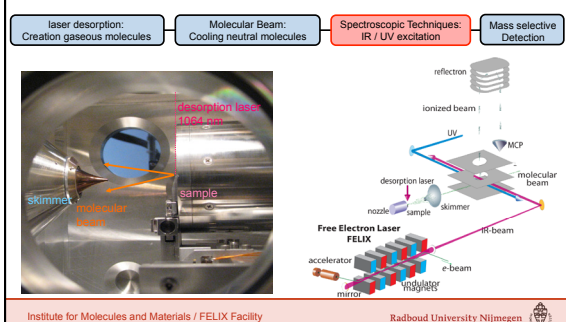
- IR-UV ion-dip spectroscopy

- provides detailed structural information / conformation selective
- direct view on hydrogen bond interactions (shift in IR frequency)

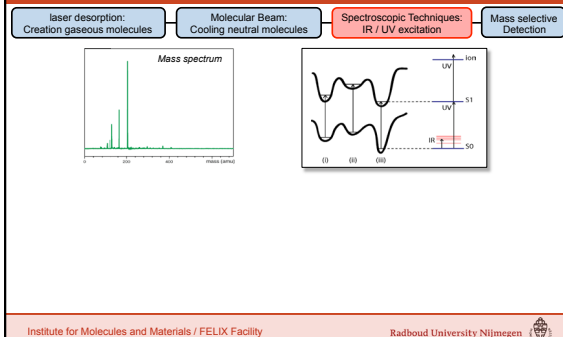
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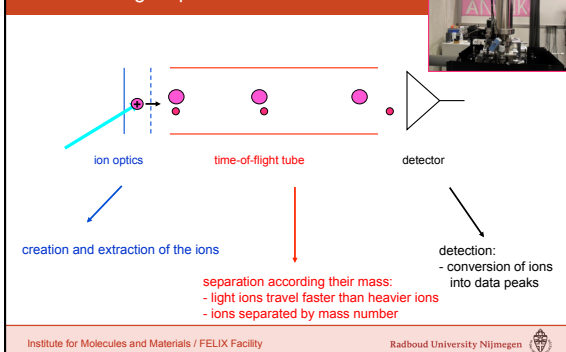
Experimental Method: Molecular Beam



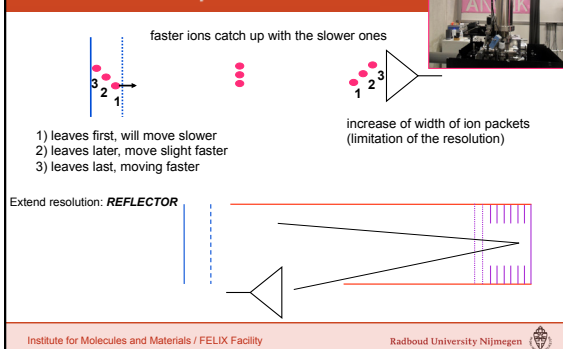
The experiment



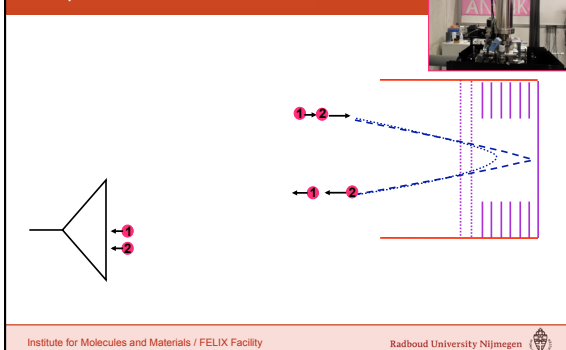
The time-of-flight spectrometer



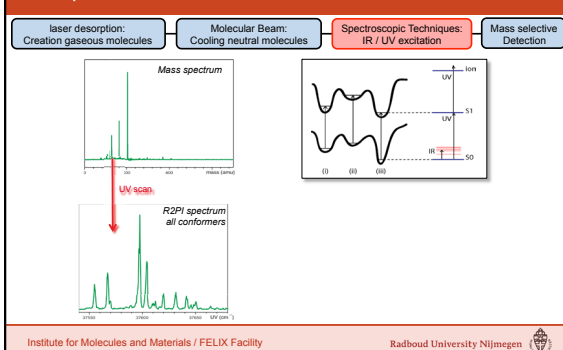
TOF-spectrometer: *spatial distribution*



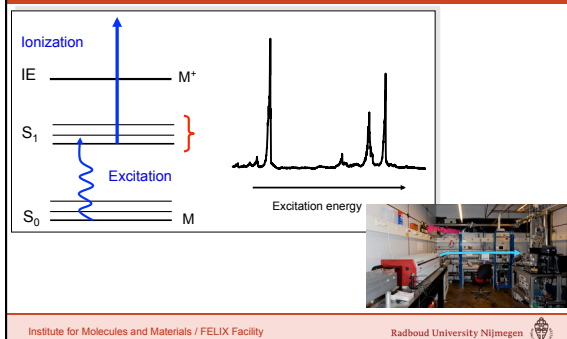
TOF-spectrometer: *the reflector*



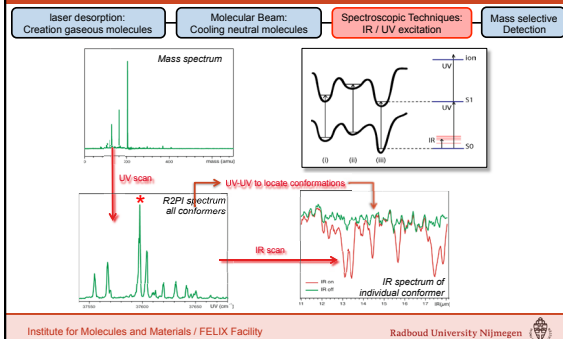
The experiment



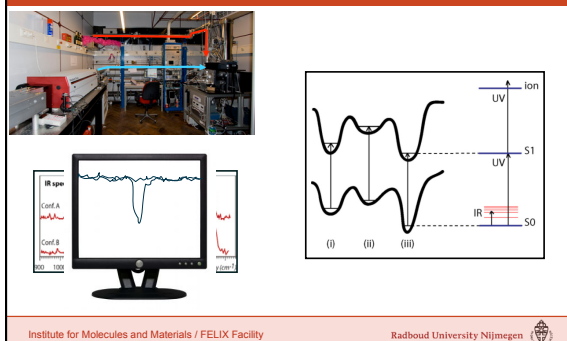
REMPI (resonant enhanced multiphoton ionization)



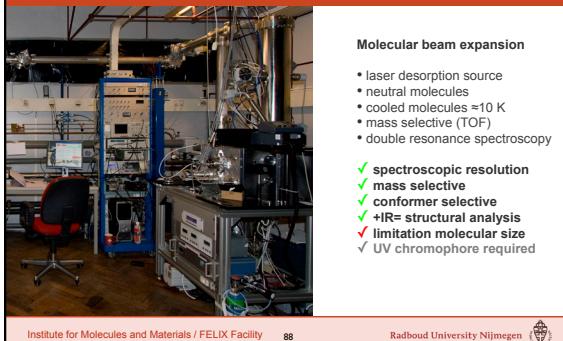
The experiment



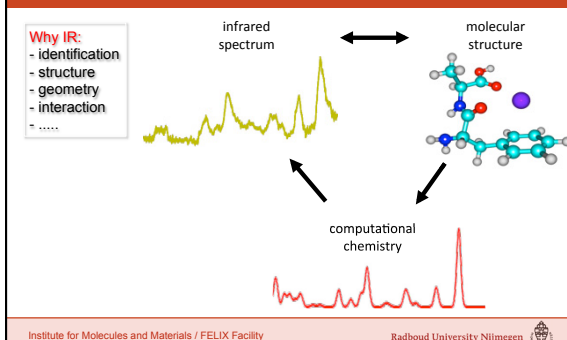
IR spectroscopy



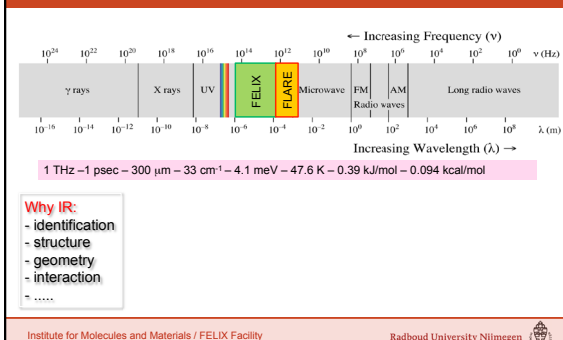
Molecular beam + laser desorption



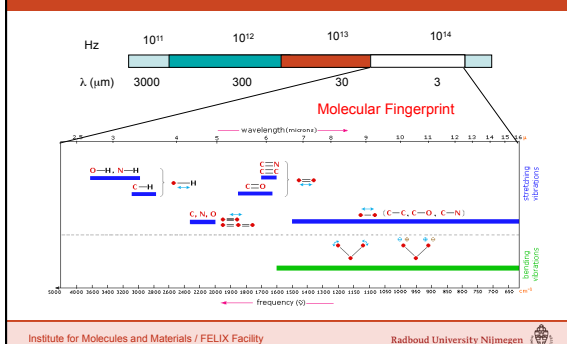
Infrared spectroscopy – molecular structure



Why is the IR / terahertz region interesting?



The fingerprint region

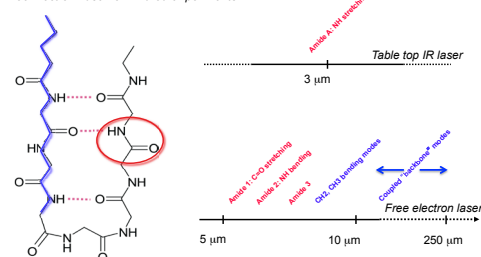


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The fingerprint of a peptide: need for FELIX

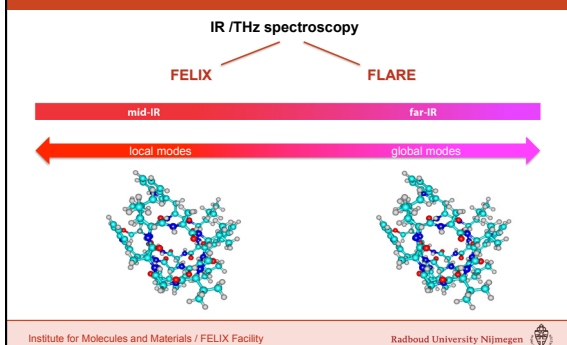
FELIX : Free Electron Laser for Infrared eXperiments



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Why is the IR / terahertz region interesting?



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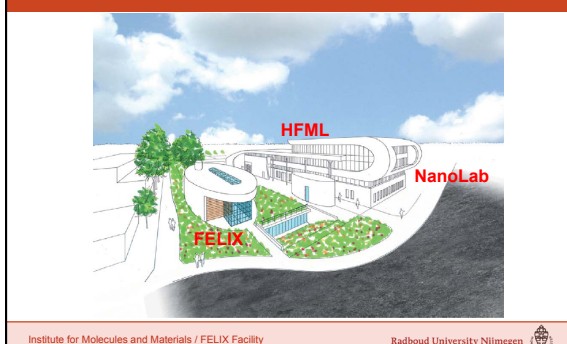
The merge: new FELIX facility Nijmegen



Institute for Molecules and Materials / FELIX Facility

Radboud University Nijmegen

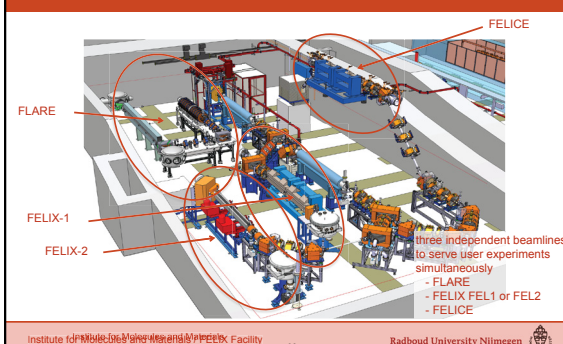
The new FELIX facility Nijmegen



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FELIX facility: Infrared Sources

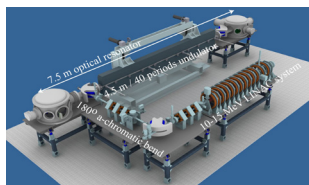


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What is a Free Electron Laser (FEL)

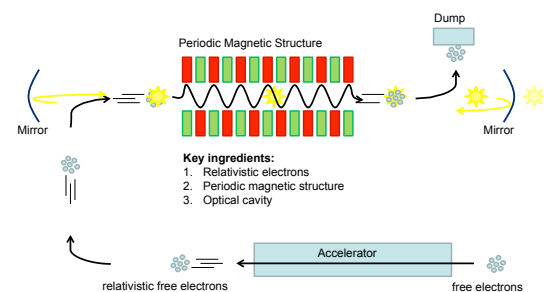
A laser light source where free electrons generate the light. In everyday-life lasers the electrons are bound in the lasing medium (gasses, dyes, solid state diodes)



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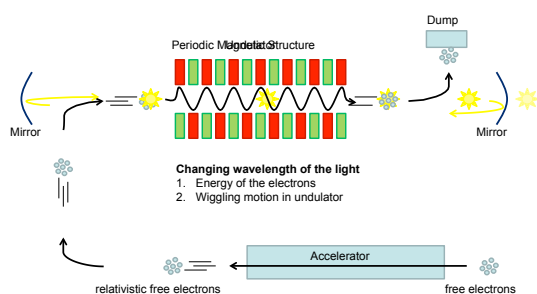
How an FEL works



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How an FEL works



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Free electron Laser for Advanced spectroscopy and high Resolution Experiments

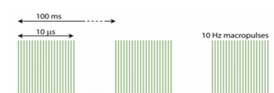


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FLARE – specifications

- Spectral range: 0.1 – 1.5 mm (0.2 – 3 THz / 6 – 100 cm^{-1})
- Repetition rate: 10 Hz
- Intense pulses of light:
 - Typically microJoules in a picosecond pulse (~100 kW)
 - Train of picosecond pulses that lasts for 10 microseconds
 - 0.5 – 2% bandwidth



- Specialty:
 - Narrow bandwidth operation (10^{-5} – 10^{-6})

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FELIX Facility: Specs of the Infrared Sources

SPECS:	FELIX:	FLARE:	FELICE:
e-beam energy	50/45 – 15 MeV	15 – 10 MeV	10/45 – 10 MeV
spectral range	2.7 – 150 micron 3500 – 66 cm^{-1} 120 – 2 THz 1500 – 8 meV	100 – 1500 micron 100 – 6 cm^{-1} 3 – 0.25 THz 12 – 0.75 meV	6 – 100 micron 1000 – 100 cm^{-1} 10 – 3 THz 100 – 12 meV
pulse structure	micro / macropulses	micro / macropulse	micro / macropulses
rep. rate	25 MHz/1 GHz@10 Hz	3 GHz/20 MHz@10 Hz	10 MHz/20 MHz@10 Hz
micropulse energy	1–20 μJ	$\approx 5 \mu\text{J}$	max. 1 mJ
macropulse energy	$\leq 100 \text{ mJ}$ @ 1 GHz	$\leq 100 \text{ mJ}$ @ 1 GHz	max. 5 J @ 1 GHz
peak power	$\leq 100 \text{ MW}$	$\leq 10 \text{ MW}$	$\leq 5 \text{ GW}$
polarisation	linear	linear	linear
spectral bandwidth (FWHM)	0.2 – 5%	$\leq 1\%$ * spectral mode $\leq 10^{-4}$	0.1 – 1%
continuous tunability	200 – 300%	7%	max. 100%

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FELIX facility @ Nijmegen: User Laboratories

User laboratory 1 – FLARE & FELIX

He-droplet machine Havenith (Bochum)	EPR spectrometer EPSRC, Aeppli, Mordin	Ultrafast laser system Kimmel & Rasing (RU)	FLARE Diagnostic Station
Cold 22-pole ion trap Schlemmer (Cologne)		Molecular beam apparatus	Metal cluster setup + Water cluster machine
earmarked for electron diffraction setup (FLARE collaboration with Prof. J. Lutten, TUE)			
multi-purpose station with optical table			

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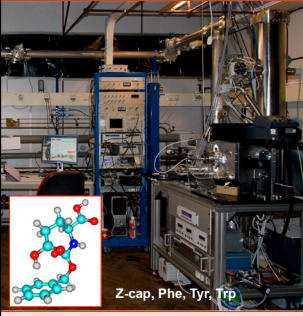
FELIX facility @ Nijmegen: User Laboratories

User laboratory 2 – FELIX & FELICE

Ultrafast laser systems	Non-linear optics laboratory
Versatile FTICR mass spectrometer	FELICE FTICR mass spectrometer
Multi- purpose ion trap	FELICE cluster apparatus
FELIX diagnostic station	

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Molecular beam + laser desorption



Molecular beam expansion

- laser desorption source
- neutral molecules
- cooled molecules ≈ 10 K
- mass selective (TOF)
- double resonance spectroscopy

- ✓ spectroscopic resolution
- ✓ mass selective
- ✓ conformer selective
- ✓ +IR= structural analysis
- ✓ limitation molecular size
- ✓ UV chromophore required

Z-cap, Phe, Tyr, Trp

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Natural amino acids

Small	Nucleophilic	Hydrophobic	Aromatic	Acidic
<chem>NC(C(=O)O)C(=O)O</chem> Glycine (Gly, G) MW: 75.07	<chem>NC(C(C)C(=O)O)C(=O)O</chem> Alanine (Ala, A) MW: 77.09	<chem>NC(C(C)C(=O)O)C(=O)O</chem> Valine (Val, V) MW: 99.09	<chem>NC(C1=CC=CC=C1C(=O)O)C(=O)O</chem> Phenylalanine (Phe, F) MW: 147.15	<chem>NC(C(C(=O)O)C(=O)O)C(=O)O</chem> Aspartic Acid (Asp, D) MW: 119.09
<chem>NC(C(C(C)C(=O)O)C(=O)O)C(=O)O</chem> Serine (Ser, S) MW: 87.08	<chem>NC(C(C(C)C(=O)O)C(=O)O)C(=O)O</chem> Leucine (Leu, L) MW: 133.17	<chem>NC(C(C(C)C(=O)O)C(=O)O)C(=O)O</chem> Isoleucine (Ile, I) MW: 132.16	<chem>NC(C1=CC=C(C=C1)C(=O)O)C(=O)O</chem> Tryptophan (Trp, W) MW: 188.17	<chem>NC(C(C(C(=O)O)C(=O)O)C(=O)O)C(=O)O</chem> Glutamic Acid (Glu, E) MW: 120.09
<chem>NC(C(C(C)C(=O)O)C(=O)O)C(=O)O</chem> Threonine (Thr, T) MW: 119.09	<chem>NC(C(C(C)C(=O)O)C(=O)O)C(=O)O</chem> Methionine (Met, M) MW: 149.15	<chem>NC(C(C(C)C(=O)O)C(=O)O)C(=O)O</chem> Proline (Pro, P) MW: 97.09	<chem>NC(C(C(C)C(=O)O)C(=O)O)C(=O)O</chem> Arginine (Arg, R) MW: 174.20	<chem>NC(C(C(C)C(=O)O)C(=O)O)C(=O)O</chem> Histidine (His, H) MW: 133.07
<chem>NC(C(C(C)C(=O)O)C(=O)O)C(=O)O</chem> Lysine (Lys, K) MW: 146.15	<chem>NC(C(C(C)C(=O)O)C(=O)O)C(=O)O</chem> Asparagine (Asn, N) MW: 132.12	<chem>NC(C(C(C)C(=O)O)C(=O)O)C(=O)O</chem> Glutamine (Gln, Q) MW: 146.15	<chem>NC(C(C(C)C(=O)O)C(=O)O)C(=O)O</chem> Tyrosine (Tyr, Y) MW: 181.19	<chem>NC(C(C(C)C(=O)O)C(=O)O)C(=O)O</chem> Cysteine (Cys, C) MW: 121.16

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VUV: 1-photon ionization

Why VUV?

Without UV-chromophore

VUV single photon ionization

355 nm frequency tripling

Xe-Ar (1:10), 200 Torr

118 nm (10.6 eV)

IR-VUV spectroscopy

Ac-Ala-Cys-NHMe

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 - Molecular beams & laser desorption

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- Free electron lasers
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 - Combination with VUV sources

Applications

- Biomolecules
 - Local environment
 - Biological environment
 - Larger and larger...
- Molecular motors: functioning of rotaxanes

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